

International Journal of Environment and Climate Change

Volume 13, Issue 8, Page 1871-1882, 2023; Article no.IJECC.101665 ISSN: 2581-8627 (Past name: British Journal of Environment & Climate Change, Past ISSN: 2231–4784)

# Seed Priming: A Key to Sustainability in Drought Stress

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#### Authors' contributions

This work was carried out in collaboration among all authors. Author SA designed the study, performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Authors SC and VH managed the analyses of the study. Author SL managed the literature searches and correspond the article. All authors read and approved the final manuscript.

#### Article Information

DOI: 10.9734/IJECC/2023/v13i82142

#### **Open Peer Review History:**

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/101665

**Review Article** 

Received: 13/04/2023 Accepted: 15/06/2023 Published: 17/06/2023

#### ABSTRACT

Abiotic stresses spot lights the field crops at all growth phases and results in significant yield losses in important crops, endangering the safety of the world's food supply. Numerous physiological, biochemical, and molecular tactics have been examined by crop researchers to battle drought stress/water limiting stress, but in the current environment, these measures are meager. It is so claimed that plants can be primed by various organic and inorganic stimulants for exceptional toughness under stressful circumstances. In order to confer tolerance, novel seed priming techniques are promising field of research in stress biology and crop stress management. Seed priming is the process of carefully hydrating seeds with germination stimulants so that pregerminative metabolic activity can continue while the radicle's emergence is halted. The terms "hydro-priming," "osmo-priming," "halo-priming," "solid matrix priming," "bio-priming," and "hormonal priming" refer to various priming techniques. It will speed up and synchronize germination, improve plant growth and stand establishment, raise stress tolerance, improve fertilizer and water use

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Int. J. Environ. Clim. Change, vol. 13, no. 8, pp. 1871-1882, 2023

efficiency, and have superior weed suppression effects. This review paper covers the physiological and biochemical changes through several seed priming techniques, Nano-priming and its significance in sustainable agriculture.

Keywords: Seed priming; water limited stress; seed priming techniques; nano priming.

#### 1. INTRODUCTION

For the vast majority of India's poor and vegetarian people, pulses are a staple food and a significant source of protein. Pulses offer the ideal combination of high biological value vegetarian protein when combined with cereals. According to the findings of household consumption surveys, there has been a fall in pulse consumption, which has increased malnutrition and decreased protein intake [1]. About 24% of the world's undernourished population still live in India [2]. According to GOI report (2016), a plunge in the country's per capita net availability of pulses from 70.3 to 29.1 g/day/capita was observed in years 1959 to 2003. With the production of pulses in the recent decades, an improvement in availability of pulses in vegetarian population in India. According to projections from the Indian Institute of Pulses Research, Kanpur, the nation's pulses demand reach 39 million tonnes by will 2050. necessitating a 2.2 percent annual growth in production [3]. According to GOI report (2016), a plunge in the country's per capita net availability of pulses from 70.3 to 29.1 g/day/capita was observed in years 1959 to 2003. With the production of pulses in the recent decades. an improvement in availability of pulses in India has noticed which was mainly due to pulse related initiatives (policies and programmes) which works in the direction of boosting the domestic production and increased imports, and is currently at 47.2 g/day/capita [4]. Pulses, as an important source of protein, constitute a basic

ingredient in the diet of vast majority of poor and vegetarian population in India. According to projections from the Indian Institute of Pulses Research, Kanpur, the nation's pulses demand reach 39 million tonnes by 2050. will necessitating a 2.2 percent annual growth in production [3]. The Economics times 2020 reported the total production of pulses for 2019-2020 was 23.02 million tones which was shown as increase of 2.76 million tonnes from the fivevear average production of 20.26 million tonnes [7]. India accounts for about 29% of the global area and 19% of the global production of pulses making it the largest producer and consumer in the world. More significantly, India is the world's largest importer and processor of pulses. In comparison to the global average of 1023 kg/ha the country's average production is roughly 841 kg/ha [5]. Agriculture is the most vulnerable sector to water constraint and is experiencing a large (40 to 60 percent) loss in output potential in rain-fed areas [6]. By 2050, it is anticipated that there will be an additional 2 billion people on the planet, which will increase the need for food to feed the expanding population [8]. However, the main problems impeding efforts to meet the world's food needs are global warming, climate change, and an ever-increasing population [9]. Abiotic stress are the atrocious challenges that come forward mainly due to unfavorable climatic change scenario, such challenges are heavy metal toxicity, drought, heat, and high soil salinity, which ultimately decreased crop output across the globe [10].

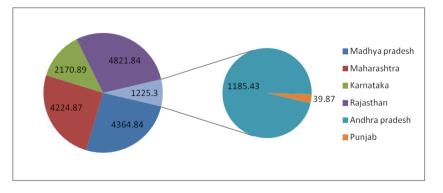


Fig. 1. State wise production of pulses (in tonnes) during 2020-21, Ministry of Agriculture & Farmers Welfare, (2021) https://pib.gov.in/Press

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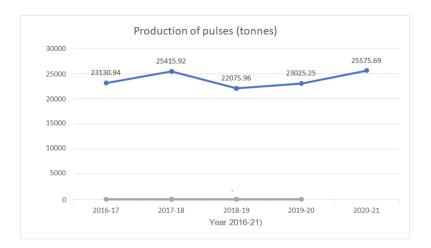


Fig. 2. State wise production of pulses (in tonnes) during 2016-21, Ministry of Agriculture & Farmers Welfare, (2021) https://pib.gov.in/Press

#### 2. EFFECT OF DROUGHT STRESS ON GROWTH AND DEVELOPMENT OF PLANT

The main issues impacting seed germination, emergence, seedling vigour, and ultimately crop output are abiotic stresses. These unfavorable significantly affect circumstances manv agricultural areas in dry and semi-arid countries. Drought can interfere with a plant's natural functioning at various phases of growth, which lowers the plant's overall production [11]. Plants adjust physiologically, biochemically, and molecularly to these conditions to some extent, but these natural adaptation techniques are insufficient to produce the desired results [12,13]. Low germination percentage and less stand establishment reduces overall arowth of seedlings by affecting their morphological (seedling length and biomass), physiological (relative water content), biochemical (amylase, protease, and lipase activities), and molecular (stress proteins, aquaporins, and dehydrins) characteristics [14,15]. Additionally, it generates reactive oxygen species (ROS) and affects the cell in two ways firstly, disrupts cell membranes which results in electrolytes leakages secondly limitation in electron transfer, quenching of excitation energy which produces  $O_2$  in PSII, reduction in  $O_2$  to  $O_2$  [16]. Reproductive phase the sensitive growth phase of plant determines yield in most crops is also affected by drought stress [17,18]. Drought stress during the reproductive period affects grain development, grain number, grain weight, and many other vields and vield-attributing features, which leads to a significant decrease in productivity and quality of final production [19,20]. Being an

agricultural nation, India needed simple, efficient, and manageable technologies to improve crop establishment under all environmental circumstances. Seed priming is a straightforward and effective wav to svnchronize seed germination, promote emergence, and establish in the farm, among other methods for increasing crop yield. By synchronizing seed germination, the seed priming approach can help farmers reduce the losses by drought stress, increase crop production, and create systemic resilience to drought stress in plants.

#### 2.1 Seed Priming

Simple and efficient hydration strategy to promote seed germination is seed priming. Seeds undergo a physiological process known as regulated hydration and drying during priming, which results in an improved and increased pregerminative metabolic process for quick germination [21]. Prior to germination, seed treatments cause a physiological state known as the "primed state," which enhances a number of cellular reactions [22]. The prepared seeds produce seedlings that have early, uniform germination and an overall improvement in its lifetime different growth characteristics might be seen [23,24]. It improves crop production, nutrient uptake, water use effectiveness, release of photo- and thermos-dormancy and activating genes/ proteins that respond to stress [25], such as late embryogenesis abundant (LEAs), which may result in the development of drought stress resistance [26,27].

#### 2.2 Techniques of Priming

Every crop needs a unique, optimized priming procedure. Optimization takes into account a

number of factors, including the amount of time needed for treatment, the priming or coating material, the vigour of the seeds, and the storage circumstances (temperature, moisture, oxygen requirement etc.)

### 2.2.1 Nano priming: A realistic seed priming technique

The rapidly expanding area of nanotechnology has several industrial applications in the fields of pharmaceuticals, food, cosmetics, electronics, textiles, energy, environmental bio-remediation etc. [28]. Currently, agriculture sector is also interested and performing wonders in applications of nanotechnology in field of seed biology [29]. Numerous researchers have been conducted extensive research on the use of nano particles for crop protection and vield enhancement against a variety of biotic and abiotic challenges [30]. To comprehend how Nanoparticles (NPs) and plants interact, many studies have been conducted and presented significant data on various research platforms. However, due to the association of NPs differing from species to species, a variety of results based on the physiological and molecular parameters of crops have been seen. In addition, NPs respond differently depending on their size, shape, manufacturing method and chemicals used [31]. Plant growth is altered when NPs penetrate the cell wall and cause a variety of morphological and physiological alterations.

### 2.2.2 Nano technology in sustainable agriculture

Nanotechnology offers enormous promise in agriculture, such as minimizing the effects of climate change and enhancing abiotic stress management measures [44]. Nano-enabled technologies have been developed to promote plant growth, such as the application of nano fertilizers through various irrigation methods (such as soil irrigation, foliar spray, and seed coating), nano-sensors to monitor real-time plant health, and genetic engineering of plants to increase defense-related phytohormones and photosynthetic efficiency. Application of nano fertilizers may have high surface area-to-volume ratio, high pollutant removal effectiveness, and efficient provision of critical nutrients for soil health over conventional fertilizers [45]. Several research have reported on the use of NPs as nano fertilizers to boost crop yield under stress [46,47]. Nutrient retention circumstances capacity (high surface area) of NPs reduces

nutrient losses and provides potential benefits to plants [48,49]. NPs as nano fertilizers improved abiotic stress tolerance in plants by enhancing plant growth, nutritional content, phytohormones, antioxidant enzymes, and photosynthetic efficiency while decreasing cellular oxidative stress. Recently, iron oxide (Fe<sub>3</sub>O<sub>4</sub>) NPs have been employed to boost crop development in both high mechanical stress-contaminated soil and drought-stressed circumstances [50,51]. Furthermore, [52] demonstrated that FeO NPs improved wheat seedling growth by reducing oxidative stress caused by Cd and Pb contamination [53]. The use of nanoparticles effectively alleviates salt stress by lowering salt content and the harmful effects associated with it. Furthermore, nano-silicon (Si) has been shown to greatly improve salt stress, seed germination, the antioxidant defence system, leaf turgor, and the carbon-assimilation process [54]. [55] recently showed that the application of cerium oxide (CeO) NPs maintained quantum vield of photosystem II and CO<sub>2</sub> assimilation via ROS scavenging, particularly hydrogen peroxide, produced by abiotic stress [56]. Titania (TiO<sub>2</sub>) nanoparticles increased catalase (CAT), glutathione peroxidase (GPOX), and superoxide dismutase (SOD) activities and reduced oxidative stress in Duckweed (Lemna minor) plants [57].

#### 2.2.3 Nano Priming under drought stress

Drought is a major environmental condition that has piqued the interest of both environmental and agricultural specialists. Limited moisture content diminishes cell size, disrupts membrane integrity, induces oxidative stress, and causes leaf senescence, all of which reduce crop output [58]. Previous research has shown that Si NPs improve plant drought tolerance. Drought resistance, for example, improved in hawthorn plants treated with SiNPs, but defense-related physiological indicators varied according to drought levels and SiNPs concentrations applied [59]. Similarly, Si NPs revealed a high potential for post-drought plant recovery in barley plants modifying morphophysiological through characteristics [60,61] discovered that Si NPs improved cucumber growth and yield under water-stressed and saline situations [85].

Under drought conditions, chitosan NPs enhanced the relative water content, photosynthetic rate, CAT, SOD activities, yield, and biomass of wheat plants [62]. Foliar application of Fe NPs has been shown to alleviate drought stress effects on safflower cultivars [63], whereas soil application of CeO NPs improved plant growth at 100mg/kg and increased photosynthetic rate by regulating water use efficiency in soybean (*Glycine max*) plants [64]. The application of silver nanoparticles minimized the harmful effects of drought stress on lentils (*Lens culinaris* Medic.) plants [65]. [66] Identified Si NPs-assisted Abscissic acid administration as an efficient drought resistance management approach in Arabidopsis thaliana.

## 2.2.4 Nano-priming stimulates seed germination and seedling growth

Seed germination is a key and sensitive stage of plant life that involves a variety of metabolic activities; overcoming environmental challenges

at this point of the life cycle is critical for later growth stages [67]. In comparison to unprimed priming seeds. can synchronize seed germination and increase seedling vigour under stress circumstances [68,69]. The effects of nano-priming on seed germination have been studied, and NPs such as zinc and zinc oxide, iron, titanium oxide, and silver NPs have been widely used in this regard [70,71]. Nano-priming improved photosynthetic parameters, maintained biochemical balance, and increased biomass production in the wheat seedlings as compared to the untreated seeds. Additionally, compared to unprimed seedlings, seed-priming wheat seeds with ZnONPs increased shoot height and rootshoot biomass in seedlings under saline stress [72].

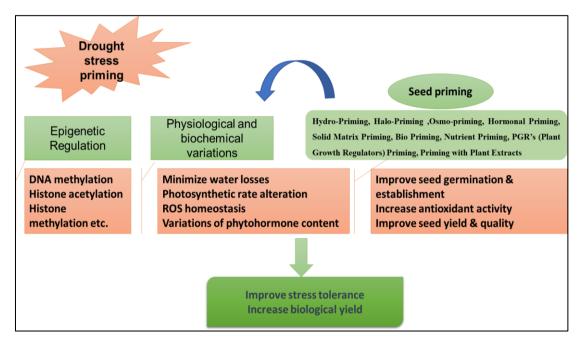


Fig. 3. A Seed priming effects under drought stress

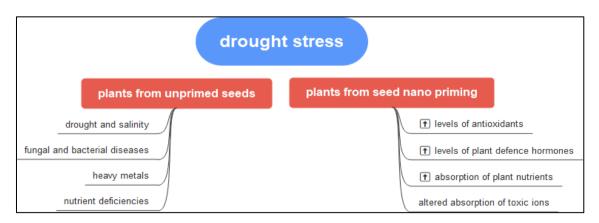


Fig. 4. Difference between primed and unprimed seeds under drought stress

S. No.	NPs agents used in studies	Effects on different crops	References
1.	Iron-oxide nanoparticles	Crop: Sorghum (Sorghum bicolor (L.) Effects: increase Seed and seedlings vigor, Biochemical activity, Biomass, and water	[32]
		content in leaves.	
2.	Zinc oxide and iron oxide nanoparticles	Crop: Wheat ( <i>Triticum aestivum</i> L.). Effects: increase Plant morphology, Biomass, Biochemical activity, Cadmium uptake, and Biofortification.	[33]
3.	Silicon nanoparticles	Crop: Wheat ( <i>Triticum aestivum</i> L.). Effects: increase Biomass, Biochemical activity, ROS levels, and Cadmium uptake.	[34]
4.	Manganese (III) oxide nanoparticles	Crop: Jalapeño ( <i>Capsicum annuum</i> L.). Effects: increase Salinity resistance, Antioxidant enzymes.	[35]
5.	Iron (II) sulfide aqua nanoparticles	Crop: Rice ( <i>Oryza sativa</i> L.). Effects: increase Seed and seedlings vigor, Plant morphology, Anti-microbial activity.	[36]
6.	Platinum nanoparticles stabilized with poly (vinylpyrroli done)	Crop: Pea ( <i>Pisum sativum</i> L.). Effects: increase Seed and seedlings vigor, Plant morphology, and decrease micro- organisms colonization (Arbuscular Mycorrhizal fungi (AMF) and rhizobia	[37]
7.	Iron nanoparticles	Crop: Wheat ( <i>Triticum aestivum</i> L.), types WL711 (low-iron genotype) and IITR26 (high- iron genotype). Effects: increase Seed and seedlings vigor, Plant morphology, and Harvest.	[38]
8.	Copper and Iron nano particles	Crop: Wheat ( <i>Triticum aestivum</i> L.) seeds of varieties galaxy-13, Pakistan-13, and NARC- 11. Effects: increase Enzymatic activity, Biochemical activity, Anti-oxidant activity, Abiotic stress resistance, and Harvest.	[39]
9.	Cobalt and Molybdenum oxide nanoparticles	Crop: Soybean seeds ( <i>Glycine max</i> (L). Merr.). Effects: increase Seed and seedlings vigor, Plant morphology, Biomass, and Enzymatic activity.	[40]
10.	Molybdenum Nanoparticles Combined with the bacteria Meso rhizobiu cicero ST-282 And Bacillus-subtilis Ch13	Crop: Chickpea ( <i>Cicer arietinum</i> L.). Effects: increase Seed and seedlings vigor, Plant morphology, Antioxidant enzymes, and Harvest.	[41]
11.	Nanoparticles of zinc, titanium, and silver	Crop: Chilli ( <i>Capsicum annuum</i> L.). Effects: increase Seed and seedlings vigor, Plant morphology, Anti-microbial activity.	[42]
12.	Copper nanoparticles	Crop: Common bean ( <i>Phaseolus vulgaris</i> L.). Effects: increase Seed and seedlings vigor, (High concentrations inhibited seed germination, independent of nanoparticle size), and Biomass.	[43]

Additionally, nano-priming with ZnO, the damaging effects of salt stress on overall growth, photosynthetic pigments, and ultra-structure of leaves were reduced. Similar to this, the use of ZnONPs as a priming factor in the Brassica napus cultivars significantly improved seedling growth under salt stress and showed an increase

in the length of shoots (9.60% and 25.63%) and roots (41.64% and 48.17%), respectively, as well as increased biomass [73].

*Festuca ovina* seeds exposed to drought stress showed increased vigour after being supplemented with AgNPs (25, 50, and 75%); length of root and shoot, their corresponding wet and drv weights, vigour index, and coefficient of the seedlings increased after nano-priming, with maximum values reported at 75% concentration of AgNPs [74]. Additionally, the addition of titanium oxide nanoparticles (TiO<sub>2</sub>NPs) to maize seeds increased the length of the seedlings' roots and their fresh and dry weights under both normal and salinity-stress circumstances [75]. In this study, it was discovered that nano-priming with TiO<sub>2</sub> was more successful at encouraging seedling growth than hydro-priming. Following TiO<sub>2</sub>NPs treatment, increased relative water content, K+ concentration, total proline content, phenolic contents enhanced seedling and growth. In a recent experiment, [76] primed rapeseeds under salt stress with cerium oxide nanoparticles (nano-ceria), which improved K+ retention and decreased Na+ accumulation, changing the Na+/K+ ratio to handle salt stress as compared to control seedlings. Some nanoparticles can be poisonous and have negative effects on many growth parameters in addition to being helpful for seedling growth, which calls for diligent research in the field of nano-priming [77]. For example, priming pearl millet (Pennisetum glaucum) seeds with 5 ml of 20 and 50 mg L<sup>1</sup> of AgNP solutions slowed the growth of the seedlings in terms of the length of their roots and shoots [78]. Additionally, French bean (Phaseolus vulgaris cv. Contender) seeds primed with two engineered nanomaterials-nano chitosan (Cs, 10%) or carbon nanotubes (CNTs, 20 g L<sup>-1</sup>) either combined with NPK-displayed alone or reduction in all seedling growth parameters as compared to the control, including the root and shoot length, fresh and dry weight, leaf area, and water content [79]. However, foliar treatment of nanomaterials at the same these dose demonstrated a different effect than seed priming.

#### 2.2.5 Nano-priming under ROS equilibrium

Nano-primed plant seeds and seedlings have an species ordered reactive oxygen (ROS) homeostasis system and antioxidant that protects seeds from oxidative damage and ensures seed longevity [80]. Under normal conditions, the use of nano-scale zero-valent iron (G-nZVI) increased ROS generation, which aided in the germination of (Oryza sativa L. cv. Gobindobhog) seeds [81]. Similarly, 25 mg L<sup>-1</sup> of nanoscale micronutrient iron (Fe<sub>2</sub>O<sub>3</sub>) increased ROS generation in Oryza sativa and Zea mays seeds to accelerate seed germination when

compared to hydro-primed seeds [82]. Under stress conditions, nano-priming establishes a generation between ROS balance and scavenging in seeds. In manganese (Mn)stressed Helianthus annuus (L.), for example, 12.5- 200 M Sulphur nanoparticles (SNPs)priming caused a decrease in total ROS contentsuperoxide radical (O<sub>2</sub>) and hydrogen peroxide  $(H_2O_2)$ , as well as increased catalase (CAT) and superoxide dismutase (SOD) antioxidant activity [83]. In another experiment, TiO<sub>2</sub> nano-priming (60 ppm) of salt-stressed maize seedlings increased SOD and CAT activities while decreasing malondialdehvde (MDA-another oxidative stress marker) content and membrane electrolyte leakage (MEL) [84]. When 0.05% chitosan nanoparticles (CsNPs) were employed for priming treatment of broad bean seeds, the activities of ROS scavenging antioxidant (POX). CAT, peroxidase enzvmesand ascorbate peroxidase (APX) increased. In contrast, 0.1% of CsNPs had a negative effect on these antioxidant enzymes [83].

These findings support the use of nanoparticles as a priming agent for purposeful priming; nevertheless, the type of nanoparticles, plant species, and growing circumstances should all be considered.

#### 3. CONCLUSION

A technology known as seed priming increases germination, promotes early flowering and maturity, increases crop resistance to abiotic challenges, and is both safe and effective for the environment. The discussion above makes it clear that various priming techniques have been researched on a variety of crops and have been found to be beneficial in terms of crop yield. It is also suggested that seed priming is a better solution against germination issues when seeds are grown in unfavorable conditions. However, there are still a number of restrictions on seed priming techniques. Not every priming technique will result in considerable germination and growth. In this aspect, choosing a certain priming technique is crucial to getting greater germination and eventual yield. Therefore, more thorough research is needed to choose appropriate priming techniques for certain crops in order to ensure a higher yield.

#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

#### REFERENCES

- 1. Shalendra KC, Gummagolmath PS, Sharma P, Patil SM. Role of pulses in the food and nutritional security in India. Journal of Food Legumes. 2013;26(3&4): 124-129.
- Kumari S, Kumar R, Chouhan S, Chaudhary PL. Influence of Various Organic Amendments on Growth and Yield Attributes of Mung Bean (*Vigna radiata* L.). International Journal of Plant & Soil Science. 2023 May 13;35(12):124-30.
- lipr, Indian Institute of Pulses Research (Indian Council of Agricultural Research) Kanpur; 2015. Available: www.iipr.res.in.
- Gol.Per Capita, Per Day Net Availability of Pulses. Accessed from 2016. Available:http://pib.nic.in/newsite/PrintRele ase.aspx?relid=137358
- 5. DES. Directorate of Economics and Statistics, Department of Agriculture Cooperation and Welfare, Ministry of Agriculture, Government of India, New Delhi 2018.
- 6. FAO. The State of Food and Agriculture 2020: Overcoming Water Challenges in Agriculture. FAO, Rome; 2020.
- Available:https://doi.org/10.4060/cb1447en
  Thonta R, Pandey MK, Kumar R, Santhoshini. Studies on correlation and path coefficient for growth and yield attributes in green gram (*Vigna radiata* L. Wilczek). Pharma Innovation 2023; 12(6):1910-1915. DOI: 10.22271/tpi.2023.v12.i6v.20691
- Zsögön A, Peres LE, Xiao Y, Yan J, Fernie AR. Enhancing crop diversity for food security in the face of climate uncertainty. The Plant Journal. 2022;109(2):402-14.
- Lowry GV, Avellan A, Gilbertson LM. Opportunities and challenges for nanotechnology in the Agri-tech Revolution. Nature Nanotechnology. 2019; 14(6):517-22.
- 10. Fahad S, Bajwa AA, Nazir U, Anjum SA, Farooq A, Zohaib A, Sadia S, Nasim W, Adkins S, Saud S, Ihsan MZ. Crop production under drought and heat stress: plant responses and management options. Frontiers in Plant Science. 2017: 1147.
- 11. Gupta NK, Gupta S, Kumar A. Effect of water stress on physiological attributes and their relationship with growth and yield of wheat cultivars at different stages. Journal

of Agronomy and Crop Science. 2001;186(1):55-62.

- 12. Singhal RK, Pandey S, Bose B. Seed priming with Mg (NO3) 2 and ZnSO4 salts triggers physio-biochemical and antioxidant defense to induce water stress adaptation in wheat (Triticum aestivum L.). Plant Stress. 2021;2:100037.
- 13. Dey P, Datta D, Pattnaik D, Dash D, Saha D, Panda D, Bhatta BB, Parida S, Mishra UN, Chauhan J, Pandey H. Physiological, biochemical, and molecular adaptation mechanisms of photosynthesis and respiration under challenging environments. InPlant perspectives to global climate changes. Academic Press. 2022;79-100.
- 14. Yigit N, Sevik H, Cetin M, Kaya N. Determination of the effect of drought stress on the seed germination in some plant species. Water stress in plants. 2016;43:62.
- Jyoti C, Kakralya BL, Singhal RK. Evaluation of morpho-physiological attributes of fenugreek (Trigonella foenum graecum) genotypes under different water regimes. International Journal of Plant Sciences (Muzaffarnagar). 2017;12(2): 271-81.
- Wang X, Liu H, Yu F, Hu B, Jia Y, Sha H, Zhao H. Differential activity of the antioxidant defence system and alterations in the accumulation of osmolyte and reactive oxygen species under drought stress and recovery in rice (*Oryza sativa* L.) tillering. Scientific reports. 2019;9(1): 1-1.
- 17. Chauhan J, Singhal RK, Kakralya BL, Kumar S, Sodani R. Evaluation of yield and yield attributes of fenugreek (Trigonella foenum graecum) genotypes under drought conditions. Int. J. Pure App. Biosci. 2017;5(3):477-84.
- Sodani R, Seema RK, Gupta S, Gupta N, Chauhan KS, Chauhan J. Performance of yield and yield attributes of ten Indian mustard (*Brassica juncea* L.) genotypes under drought stress. Int. J Pure App. Biosci. 2017;5(3):467-76.
- 19. Liu X, Pan Y, Zhu X, Yang T, Bai J, Sun Z. Drought evolution and its impact on the crop yield in the North China Plain. Journal of hydrology. 2018;564:984-96.
- 20. Kuwayama Y, Thompson A, Bernknopf R, Zaitchik B, Vail P. Estimating the impact of drought on agriculture using the US Drought Monitor. American Journal of

Agricultural Economics. 2019;101(1): 193-210.

- 21. Dawood MG. Stimulating plant tolerance against abiotic stress through seed priming. Advances in seed priming. 2018:147-83.
- Wojtyla Ł, Lechowska K, Kubala S, Garnczarska M. Molecular processes induced in primed seeds—increasing the potential to stabilize crop yields under drought conditions. Journal of Plant Physiology. 2016;203:116-26.
- 23. Huang L, Zhang L, Zeng R, Wang X, Zhang H, Wang L, Liu S, Wang X, Chen T. Brassinosteroid priming improves peanut drought tolerance via eliminating inhibition on genes in photosynthesis and hormone signaling. Genes. 2020;11(8):919.
- 24. Abbasi Khalaki M, Moameri M, Asgari Lajayer B, Astatkie T. Influence of nanopriming on seed germination and plant growth of forage and medicinal plants. Plant growth regulation. 2021;93:13-28.
- 25. Dutta P. Seed priming: new vistas and contemporary perspectives. Advances in seed priming. 2018:3-22.
- 26. Sen A, Puthur JT. Halo and UV-B priming influences various physiological and importantly yield parameters of Oryza sativa var. Vyttila 6. New Zealand Journal of Crop and Horticultural Science. 2021; 49(1):1-6.
- 27. Thomas TT, Puthur JT. UV-B priming enhances specific secondary metabolites in *Oryza sativa* (L.) empowering to encounter diverse abiotic stresses. Plant Growth Regulation. 2020;92:169-80.
- Moll J, Okupnik A, Gogos A, Knauer K, Bucheli TD, Van Der Heijden MG, Widmer F. Effects of titanium dioxide nanoparticles on red clover and its rhizobial symbiont. PloS one. 2016;11(5):e0155111.
- 29. Banik S, Pérez-de-Luque A. In vitro effects of copper nanoparticles on plant pathogens, beneficial microbes and crop plants. Spanish Journal of Agricultural Research. 2017;15(2):e1005- e1005.
- Li J, Hu J, Ma C, Wang Y, Wu C, Huang J, Xing B. Uptake, translocation and physiological effects of magnetic iron oxide (γ-Fe2O3) nanoparticles in corn (*Zea mays* L.). Chemosphere. 2016; 159:326-34.
- 31. Rastogi A, Zivcak M, Sytar O, Kalaji HM, He X, Mbarki S, Brestic M. Impact of metal and metal oxide nanoparticles on plant: a critical review, Front. Chem. 2017;5(78).

- Maswada HF, Djanaguiraman M, Prasad PV. Seed treatment with nano-iron (III) oxide enhances germination, seeding growth and salinity tolerance of sorghum. Journal of Agronomy and Crop Science. 2018;204(6):577-87.
- 33. Rizwan M, Ali S, Ali B, Adrees M, Arshad M, Hussain A, ur Rehman MZ, Waris AA. Zinc and iron oxide nanoparticles improved the plant growth and reduced the oxidative stress and cadmium concentration in wheat. Chemosphere. 2019;214:269-77.
- 34. Hussain A, Rizwan M, Ali Q, Ali S. Seed priming with silicon nanoparticles improved the biomass and yield while reduced the oxidative stress and cadmium concentration in wheat grains. Environmental Science and Pollution Research. 2019:26:7579-88.
- 35. Ye Y, Cota-Ruiz K, Hernandez-Viezcas JA, Valdes C, Medina-Velo IA, Turley RS, Peralta-Videa JR, Gardea-Torresdey JL. Manganese nanoparticles control salinitymodulated molecular responses in Capsicum annuum L. through priming: A sustainable approach for agriculture. ACS Sustainable Chemistry & Engineering. 2020;8(3):1427-36.
- 36. Ahuja R, Sidhu A, Bala A. Synthesis and evaluation of iron (ii) sulfide aqua nanoparticles (FeS-NPs) against Fusarium verticillioides causing sheath rot and seed discoloration of rice. European Journal of Plant Pathology. 2019;155:163-71.
- Rahman MS, Chakraborty A, Mazumdar S, Nandi NC, Bhuiyan MN, Alauddin SM, Khan IA, Hossain MJ. Effects of poly (vinylpyrrolidone) protected platinum nanoparticles on seed germination and growth performance of Pisum sativum. Nano-Structures & Nano-Objects. 2020; 21:100408.
- Sundaria N, Singh M, Upreti P, Chauhan RP, Jaiswal JP, Kumar A. Seed priming with iron oxide nanoparticles triggers iron acquisition and biofortification in wheat (Triticum aestivum L.) grains. Journal of Plant Growth Regulation. 2019;38:122-31.
- Yasmeen F, Raja NI, Razzaq A, Komatsu S. Proteomic and physiological analyses of wheat seeds exposed to copper and iron nanoparticles. Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics. 2017;1865(1):28-42.
- 40. Chau NH, Doan QH, Chu TH, Nguyen TT, Dao Trong H, Ngo QB. Effects of different nanoscale microelement-

containing formulations for presowing seed treatment on growth of soybean seedlings. Journal of Chemistry;2019.

- 41. Shcherbakova EN, Shcherbakov AV, Andronov EE, Gonchar LN, Kalenskaya SM, Chebotar VK. Combined pre-seed treatment with microbial inoculants and Mo nanoparticles changes composition of root exudates and rhizosphere microbiome structure of chickpea (*Cicer arietinum* L.) plants. Symbiosis. 2017;73:57-69.
- 42. Kumar GD, Raja K, Natarajan N, Govindaraju K, Subramanian KS. Invigouration treatment of metal and metal oxide nanoparticles for improving the seed quality of aged chilli seeds (Capsicum annum L.). Materials Chemistry and Physics. 2020;242:122492.
- 43. Duran NM, Savassa SM, Lima RG, de Almeida E, Linhares FS, van Gestel CA, Pereira de Carvalho HW. X-rav spectroscopy uncovering the effects of Cu based nanoparticle concentration and structure on Phaseolus vulgaris germination and seedling development. Journal of Agricultural and Food Chemistry. 2017;65(36):7874-84.
- Mahakham W, Theerakulpisut P, Maensiri 44. Phumying S, Sarmah AK. S, benign synthesis Environmentally of phytochemicals-capped gold nanoparticles as nanopriming agent for promoting maize seed germination. Science of the Total Environment. 2016;573: 1089-102.
- Tripathi DK, Singh VP, Prasad SM, Chauhan DK, Dubey NK. Silicon nanoparticles (SiNp) alleviate chromium (VI) phytotoxicity in *Pisum sativum* (L.) seedlings. Plant Physiology and Biochemistry. 2015;96:189-98.
- 46. Ahmed T, Shahid M, Noman M, Niazi MB, Mahmood F, Manzoor I, Zhang Y, Li B, Yang Y, Yan C, Chen J. Silver nanoparticles synthesized by using Bacillus cereus SZT1 ameliorated the damage of bacterial leaf blight pathogen in rice. Pathogens. 2020;9(3):160.
- 47. Ahmed T, Noman M, Manzoor N, Shahid M, Abdullah M, Ali L, Wang G, Hashem A, Al-Arjani AB, Alqarawi AA, Abd\_Allah EF. Nanoparticle-based amelioration of drought stress and cadmium toxicity in rice via triggering the stress responsive genetic mechanisms and nutrient acquisition. Ecotoxicology and Environmental Safety. 202;209:111829.

- Wang P, Lombi E, Zhao FJ, Kopittke PM. Nanotechnology: a new opportunity in plant sciences. Trends in plant science. 2016;21(8):699-712.
- 49. Ahmed T, Noman M, Rizwan M, Ali S, Shahid MS, Li B. Recent progress on the heavy metals ameliorating potential of engineered nanomaterials in rice paddy: a comprehensive outlook on global food safety with nanotoxicitiy issues. Critical Reviews in Food Science and Nutrition. 2021;1-5.
- Hussain A, Ali S, Rizwan M, ur Rehman MZ, Qayyum MF, Wang H, Rinklebe J. Responses of wheat (Triticum aestivum) plants grown in a Cd contaminated soil to the application of iron oxide nanoparticles. Ecotoxicology and environmental safety. 2019;173:156-64.
- 51. Adrees M, Khan ZS, Ali S, Hafeez M, Khalid S. ur Rehman MZ, Hussain A, Hussain K, Shahid Chatha SA, Rizwan M. Simultaneous mitigation of cadmium and drought stress in wheat by soil application of iron nanoparticles. Chemosphere. 2020;238:124681.
- 52. Konate A, He X, Zhang Z, Ma Y, Zhang P, Alugongo GM, Rui Y. Magnetic (Fe3O4) nanoparticles reduce heavy metals uptake and mitigate their toxicity in wheat seedling. Sustainability. 2017;9(5):790.
- 53. López-Luna J, Silva-Silva MJ, Martinez-Vargas S, Mijangos-Ricardez OF, González-Chávez MC, Solís-Domínguez FA, Cuevas-Díaz MC. Magnetite nanoparticle (NP) uptake by wheat plants and its effect on cadmium and chromium toxicological behavior. Science of the Total Environment. 2016;565:941-50.
- 54. Haghighi M, Pessarakli M. Influence of silicon and nano-silicon on salinity tolerance of cherry tomatoes (*Solanum lycopersicum* L.) at early growth stage. Scientia Horticulturae. 2013;161:111-7.
- 55. Wu H, Tito N, Giraldo JP. Anionic cerium oxide nanoparticles protect plant photosynthesis from abiotic stress by scavenging reactive oxygen species. ACS nano. 2017;11(11):11283-97.
- 56. Horie M, Nishio K, Kato H, Fujita K, Endoh S, Nakamura A, Miyauchi A, Kinugasa S, Yamamoto K, Niki E, Yoshida Y. Cellular responses induced by cerium oxide nanoparticles: induction of intracellular calcium level and oxidative stress on culture cells. The journal of biochemistry. 2011;150(4):461-71.

- 57. Song G, Gao Y, Wu H, Hou W, Zhang C, Ma H. Physiological effect of anatase TiO2 nanoparticles on Lemna minor. Environmental Toxicology and Chemistry. 2012;31(9):2147-52.
- 58. Tiwari S, Lata C, Chauhan PS, Nautiyal CS. Pseudomonas putida attunes morphophysiological, biochemical and molecular responses in Cicer arietinum L. during drought stress and recovery. Plant Physiology and Biochemistry. 2016: 99:108-17.
- 59. Ashkavand P, Tabari M, Zarafshar M, Tomásková I, Struve D. Effect of SiO2 nanoparticles on drought resistance in hawthorn seedlings. Leśne Prace Badawcze. 2015;76(4).
- Ghorbanpour M, Mohammadi H, Kariman K. Nanosilicon-based recovery of barley (Hordeum vulgare) plants subjected to drought stress. Environmental Science: Nano. 2020;7(2):443-61.
- 61. Alsaeedi A, El-Ramady H, Alshaal T, El-Garawany M, Elhawat N, Al-Otaibi A. Silica nanoparticles boost growth and productivity of cucumber under water deficit and salinity stresses by balancing nutrients uptake. Plant Physiology and Biochemistry. 2019;139:1-0.
- Behboudi F, Tahmasebi-Sarvestani Z, Kassaee MZ, Modarres-Sanavy SA, Sorooshzadeh A, Mokhtassi-Bidgoli A. Evaluation of chitosan nanoparticles effects with two application methods on wheat under drought stress. Journal of Plant Nutrition. 2019;42(13):1439-51.
- Davar ZF, Roozbahani A, Hosnamidi A. Evaluation the effect of water stress and foliar application of Fe nanoparticles on yield, yield components and oil percentage of safflower (*Carthamus tinctorious* L.). Int. J. Adv. Biol. Biomed. 16:1150–1159.
- Cao Z, Stowers C, Rossi L, Zhang W, Lombardini L, Ma X. Physiological effects of cerium oxide nanoparticles on the photosynthesis and water use efficiency of soybean (*Glycine max* (L.) Merr.). Environmental Science: Nano. 2017;4(5): 1086-94.
- 65. Das A, Das B. Nanotechnology a potential tool to mitigate abiotic stress in crop plants. Abiotic and Biotic Stress in Plants; 2019.
- 66. Sun D, Hussain HI, Yi Z, Rookes JE, Kong L, Cahill DM. Delivery of abscisic acid to plants using glutathione responsive mesoporous silica nanoparticles. Journal

of Nanoscience and Nanotechnology. 2018;18(3):1615-25.

- 67. Abbasi Khalaki M, Ghorbani A, Dadjou F. Influence of nano-priming on Festuca ovina seed germination and early seedling traits under drought stress, in laboratory condition. Ecopersia. 2019;7(3):133-9.
- Sharifi RS, Khavazi K. Effects of seed priming with Plant Growth Promoting Rhizobacteria (PGPR) on yield and yield attributes of maize (Zea mays L.) hybrids. Journal of Food, Agriculture & Environment. 2011;9(3/4 part 1):496-500.
- 69. Abbasi Khalaki M, Ghorbani Á, Dadjou F. Influence of nano-priming on Festuca ovina seed germination and early seedling traits under drought stress, in laboratory condition. Ecopersia. 2019;7(3):133-9.
- 70. Panyuta O, Belava V, Fomaidi S, Kalinichenko O, Volkogon M, Taran N. The effect of pre-sowing seed treatment with metal nanoparticles on the formation of the defensive reaction of wheat seedlings infected with the eyespot causal agent. Nanoscale research letters. 2016;11:1-5.
- 71. El-Badri AM, Batool M, Mohamed IA, Khatab A, Sherif A, Wang Z, Salah A, Nishawy E, Ayaad M, Kuai J, Wang B. Modulation of salinity impact on early seedling nano-priming stage via application of zinc oxide on rapeseed (Brassica napus L.). Plant Physiology and Biochemistry. 2021 Sep 1:166:376-92. Abou-Zeid HM, Ismail GS, Abdel-Latif SA. Influence of seed priming with ZnO nanoparticles on the salt-induced damages in wheat (Triticum aestivum L.) plants. Journal of Plant Nutrition. 2021;44(5) :629-43.
- 72. El-Badri AM, Batool M, Mohamed IA, Khatab A, Sherif A, Wang Z, Salah A, Nishawy E, Ayaad M, Kuai J, Wang B. Modulation of salinity impact on early seedling stage via nano-priming application of zinc oxide on rapeseed (*Brassica napus* L.). Plant Physiology and Biochemistry. 2021;166:376-92.
- 73. Abbasi Khalaki M, Ghorbani A, Dadjou F. Influence of nano-priming on Festuca ovina seed germination and early seedling traits under drought stress, in laboratory condition. Ecopersia. 2019;7(3):133-139.
- 74. Shah T, Latif S, Saeed F, Ali I, Ullah S, Alsahli AA, Jan S, Ahmad P. Seed priming with titanium dioxide nanoparticles enhances seed vigor, leaf water status, and antioxidant enzyme activities in maize

(*Zea mays* L.) under salinity stress. Journal of King Saud University-Science. 2021; 33(1):101207.

- 75. Khan MN, Li Y, Khan Z, Chen L, Liu J, Hu J, Wu H, Li Z. Nanoceria seed priming enhanced salt tolerance in rapeseed through modulating ROS homeostasis and α-amylase activities. Journal of Nanobiotechnology. 2021;19(1):1-9.
- 76. Singh S, Tripathi DK, Dubey NK, Chauhan DK. Effects of nano-materials on seed germination and seedling growth: striking the slight balance between the concepts and controversies. Materials Focus. 2016; 5(3):195-201.
- 77. Parveen A, Rao S. Effect of nanosilver on seed germination and seedling growth in Pennisetum glaucum. Journal of Cluster Science. 2015;26:693-701.
- 78. Abdel-Aziz HM, Hasaneen MN, Omer AM. Impact of engineered nanomaterials either alone or loaded with NPK on growth and productivity of French bean plants: Seed priming vs foliar application. South African Journal of Botany. 2019;125:102-8.
- 79. Pawar VA, Laware SL. Seed priming a critical review. Int. J. Sci. Res. Biol. Sci. 2018;5:94-101.
- Guha T, Gopal G, Das H, Mukherjee A, Kundu R. Nanopriming with zero-valent iron synthesized using pomegranate peel waste: A "green" approach for yield enhancement in Oryza sativa L. cv. Gonindobhog. Plant Physiology and Biochemistry. 2021;163:261-75.

- Prerna DI. Govindaraiu K. Tamilselvan S. 81. Kannan M, Vasantharaja R, Chaturvedi S, Influence of nanoscale Shkolnik D. micro-nutrient α-Fe2O3 on seed germination. growth. seedlina translocation, physiological effects and yield of rice (Oryza sativa) and maize (Zea mays). Plant Physiology and Biochemistry. 2021; 162:564-80.
- Ragab G, Saad-Allah K. Seed priming with greenly synthesized sulfur nanoparticles enhances antioxidative defense machinery and restricts oxidative injury under manganese stress in *Helianthus annuus* (L.) seedlings. Journal of Plant Growth Regulation. 2021;40:1894-902.
- 83. Shah T, Latif S, Saeed F, Ali I, Ullah S, Alsahli AA, Jan S, Ahmad P. Seed priming with titanium dioxide nanoparticles enhances seed vigor, leaf water status, and antioxidant enzyme activities in maize (*Zea mays* L.) under salinity stress. Journal of King Saud University-Science. 20211; 33(1):101207.
- 84. Abdel-Aziz HM, Hasaneen MN, Omer AM. Impact of engineered nanomaterials either alone or loaded with NPK on growth and productivity of French bean plants: Seed priming vs foliar application. South African Journal of Botany. 2019;125:102-8.
- 85. Chouhan S, Kumari S, Kumar R, Chaudhary PL. Climate Resilient Water Management for Sustainable Agriculture. International Journal of Environment and Climate Change. 2023;13(7):411-26.

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Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/101665