



Influence of Various Bacterial and Fungal Strains on Fruit Cuttings: A Review

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

Microbes, especially bacteria and fungi, play a significant role in the plant's multiplication outcome as well as in regulating the success of economic yield. There are some beneficial interactions made by the various fungi and bacteria, such as *Pseudomonas aeruginosa*, *Bacillus subtilis*, and endophytic fungi, which may be responsible for plant vigor, growth, shoot and root development. Bacterial and fungal strains engaged in the uptake, utilization and nutrient transportation in the earth's pores influence the distribution of essential vitamins and minerals for plant development. We may conclude that the influence of various bacterial and fungal strains on fruit cuttings is a complex and multifaceted issue. It is clear that advanced research is obligatory to better understand the precise interactions between different strains and fruit cuttings, as well as to develop strategies for managing and justifying the potential negative impacts. By achieving a deeper and clear understanding of these interactions, we can work towards optimizing health and productivity.

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1. INTRODUCTION

The influence of bacterial and fungal strains on plant propagation is a significant aspect of agriculture. Bacteria and fungi play a major role in the growth and development of plants. They also affect the overall health, nutrient absorption capacity, and disease resistance of plants. Understanding the interactions between these bacteria and plants is essential to increasing crop yields, advancing sustainable farming practices, and developing improved propagation techniques. Bacterial and fungal strains significantly affect plant proliferation through a variety of methods. Some strains have mutualistic relationships with plant roots, such as mycorrhizal fungi, which facilitate symbiotic plant uptake of nutrients, particularly phosphorus and nitrogen [1]. The general health and vigor of the plants are improved by these beneficial interactions, and this in turn influences how well the plants multiply. However, some strains of bacteria and fungi may be detrimental to plant growth. Pathogenic bacteria and fungi can cause illnesses in plants, causing growth inhibition, reduced yields, or even plant death. It is necessary to comprehend the principles behind pathogen invasion and the evolution of disease resistance in plants in order to propagate healthy crops successfully [2]. Furthermore, Bacterial and fungal strains involved in the breakdown of organic matter or nutrient cycling in the soil, have a significant impact on availability of nutrients meant for plant growth [3]. Besides this, several bacterial and fungal strains not only directly affect plant health (rooting process) and nutrient availability, but they also promote the development of advantageous microorganisms in the rhizosphere or the soil that surrounds plant roots. This may enhance a plant's overall resilience and capacity to tolerate environmental stressors, which may impact the plant's capacity to spread.

Understanding how different strains of bacteria and fungi affect plant growth is essential to developing sustainable agricultural methods. Research in this field aims to shed light on the complex relationships that exist between microorganisms and plants in order to optimize beneficial interactions and minimize the detrimental impacts of pathogenic strains. By creating techniques for multiplication and promoting the use of helpful microbes,

horticulturists and farmers can enhance crop productivity, reduce reliance on chemical inputs, and preserve the long-term health of agricultural ecosystems [4]. This review paper aims at compilation and description of the effects of various Bacterial and fungal strains on fruit cuttings.

2. BACTERIAL STRAINS AND THEIR IMPACT

Different strains of bacteria have significant effects on human health, the environment, and a variety of industrial sectors, including agriculture (plants) [5]. Some of that are described as under-

2.1 *Bacillus subtilis*

Bacillus subtilis is a typical aerobic Gram-positive soil bacterium that releases a large amount of enzymes and degrades a variety of substrates so that it may survive under ever changing environments [6]. *Bacillus subtilis* is commonly used in agriculture to treat a wide range of plant diseases. It produces antibacterial substances and catalysts that inhibit the emergence of harmful fungi and bacteria. However, its direct application in fruit cutting, which often involves manually splitting or cutting fruits, is uncommon.

2.1.1 Impact of *Bacillus subtilis* on plant growth advancement

Bacillus subtilis, an effective soil bacterium, encourages plant growth in fruit cuttings by improving access to nutrients and boosting systemic resistance. Its ability to produce growth-promoting compounds while inhibiting harmful pathogens encourages root development, leading in greater plant vigour and fruit yield [7].

2.2 *Pseudomonas aeruginosa*

Pseudomonas is a genus of rod-shaped, spore-free Gram-negative bacteria. *Pseudomonas* species are known to generate a variety of plant growth-promoting compounds, including indole acetic acid (IAA) and siderophores, which can have a good impact on roots and plant development [8].

Certain strains of *P. aeruginosa* that have the capacity to shield plants from harmful microbes. In addition to combating plant diseases, these

strains generate antimicrobial substances that protect cuttings of plants from infections and encourage root development [9].

2.2.1 Phosphate solubilization

It is commonly recognised that some *Pseudomonas* strains can solubilize phosphate, making it more available to plants. More nutrient availability can help with root production and overall plant growth [10].

2.3 *Lactobacillus plantarum*

Lactobacillus plantarum is one type of lactic acid bacterium. It is known to have relatively little effect on plant cuttings. The majority of research on *Lactobacillus plantarum* in agriculture has been on the potential advantages of utilizing rhizobacteria rather than cuttings to encourage plant growth (PGPR) [11].

2.3.1 Stress tolerance

Studies have been conducted to determine whether lactic acid bacteria, including *Lactobacillus plantarum*, can help plants become more resilient to stress. Plant cuttings treated during root development may have stronger defenses against environmental stressors [12].

The particular plant species, the surrounding environment, and the general health of the

cuttings all affect how effective *Lactobacillus plantarum* [13]. To completely comprehend the potential mechanisms behind the relationship between *Lactobacillus plantarum* and plants, more investigation is also required.

2.4 *Lactobacillus casei*

The following outcomes were recorded when applying *Lactobacillus casei* to plant cuttings:

2.4.1 Enhanced root development

Plant roots grow more quickly when *Lactobacillus casei* is present. It can promote the creation of new roots on cuttings, improving the overall health and growth of the plant [14].

2.4.2 Disease resistance

Studies have demonstrated that the antibacterial qualities of *Lactobacillus casei* may aid in shielding plant cuttings from harmful infections and illnesses. This may make the plants more robust and resilient [15].

2.4.3 Increased nutrient uptake

Lactobacillus casei may boost the quantity of nutrients that are accessible to soil-dwelling plant roots. This could lead to improved nutrient absorption and general plant health [16].

Table 1. Fruit growing and the roles played by helpful bacteria

Fruit species	Application effects	References
Banana	Increased height, width, and quantity of leaves in Cavendish banana plants. Fruit development and cluster weight were found to be statistically significantly affected.	Firettiet al. [19]
Cherry	Fruit weight variation and increased yield were noted.	Kurlus et al. [20]
Pear	Increased shoot length and production of Le Conte pear.	Khedr [21]
Strawberry	Bacteria applied at 60 mM/L NaCl concentration showed the greatest therapeutic efficacy and good defense against the stress caused by saltwater.	Nitnavare et al. [22]
Sourcherry	In sour cherry, bacteria treatment enhanced shoot length, leaf area, and yield.	Yaseen and AL-Zubaydi [23]
Blueberry	Bacteria (<i>Bacillus amyloliquefaciens</i> JC65 and JC65) enhanced the net photosynthesis rate and plant height. Grafted plants produced 14.56% more fruits of a higher grade than the control.	Yu et al. [24]
Blackberry	<i>Bacillus</i> genus affected the growth of blackberries in terms of more branches, flowers per cluster and total branches.	Robledo Buriticá et al. [25]

2.5 *Escherichia Coli*

Escherichia coli is usually associated with food-borne illness and contamination, however not all types are hazardous [17]. Specific types of *E. coli* can have an advantageous impact while cutting fruit. To begin, some not transmissible strains of *E. coli* release antibiotic compounds that can limit the growth of hazardous bacteria, improving the general safety of the fruits. Furthermore, these microorganisms can help with the inherent fermentation process, resulting in the generation of naturally occurring acids that function as natural additives, increasing the potential for preservation of cut fruits. Furthermore, *E. coli* can help promote the turnover of nutrients by digesting breakdown organic compounds during recycling, which can then be used to sustainably grow agriculture [18]. According to research, certain strains of *E. coli* could be utilized to enhance fruit consistency and flavour through fermentation procedures. However, it is critical to emphasize that stringent quality assurance procedures must be in place to guarantee that only not transmissible genotypes exist during the fruit-cutting process in order to avoid any health hazards. Overall, recognising the advantages of specific strains of *E. coli* could give rise to novel ways to fruit processing, promoting both safety and efficiency.

3. FUNGAL STRAINS AND THEIR INFLUENCE

Fungus strains impact plant proliferation in both helpful and harmful aspects.

3.1 Pathogenic Fungi

Some fungus strains can infect plants and cause illnesses, which lowers the chances of successful replication. These fungi have the ability to infect plant tissues, stunt growth, and ultimately kill the plant [26].

An infestation of infectious fungus in fruit cuttings can harm both the nutritional content as well as the security of the harvest. To begin, pathogenic moulds such as *Botrytis cinerea* or *Monilinia fructicola* can cause obvious rot and spoilage of fruits, resulting in considerable economic losses in the fruit sector. These fungi thrive in damp settings, therefore sliced fruits are especially vulnerable to their proliferation [27]. Furthermore, certain pathogenic fungi create mycotoxins, which can be harmful if ingested,

demanding strict surveillance and standard control procedures throughout the fruit cutting process. The growth of mould on cut layers can detract from the entire visual attractiveness of the fruit and negatively affect customer impressions [28]. Furthermore, pathogenic fungus can accelerate deterioration, shortening the life expectancy of cut fruits and raising the risk of cross-contamination. Maintaining strict hygiene procedures, adequate storage conditions, and the use of antifungal agents are critical in reducing the detrimental impact of pathogenic fungus on fruit cuttings and assuring the safety of the finished product.

3.2 *Trichoderma* spp.

Trichoderma, a genus of soil-borne fungus, has received a lot of attention for its wide range of interactions with plants, including mycoparasitism and symbiosis. Besides this, some species of *Trichoderma* are also helpful as biopesticides. It promotes sustainable and ecologically friendly methods by lowering the need for artificial fertilizers and pesticides [29]. These are helpful for improvement in pomegranate cutting establishments due to their growth-promoting and biocontrol properties. It is also established for the bio control of soil borne infections, *Trichoderma* species have been extensively employed [25]. These molds have the capacity to form symbiotic connections with plant roots in addition to encouraging plant growth and enhancing the plant's resistance to stress [30]. Because of these interactions, bioactive substances like enzymes and antibiotics are produced which improve plant health and reduce the frequency of disease [31].

3.3 *Trichoderma*-Induced Endophytic Microbiota Stimulates Plant Immunity Through a Synergistic Mechanism

3.3.1 Comparative functions of *Trichoderma*

These are saprophytic fungi that develop quickly on mycelial filaments and have a high degree of environmental adaptation. By becoming entangled in the invasive section of the roots of pathogenic fungi, it can prevent their spread. Moreover, it might take up the nutrients needed for the disease-causing fungi to grow, creating a nutritional deficient environment that prevents the fungi from multiplying and growing [32] (Fig. 1A).

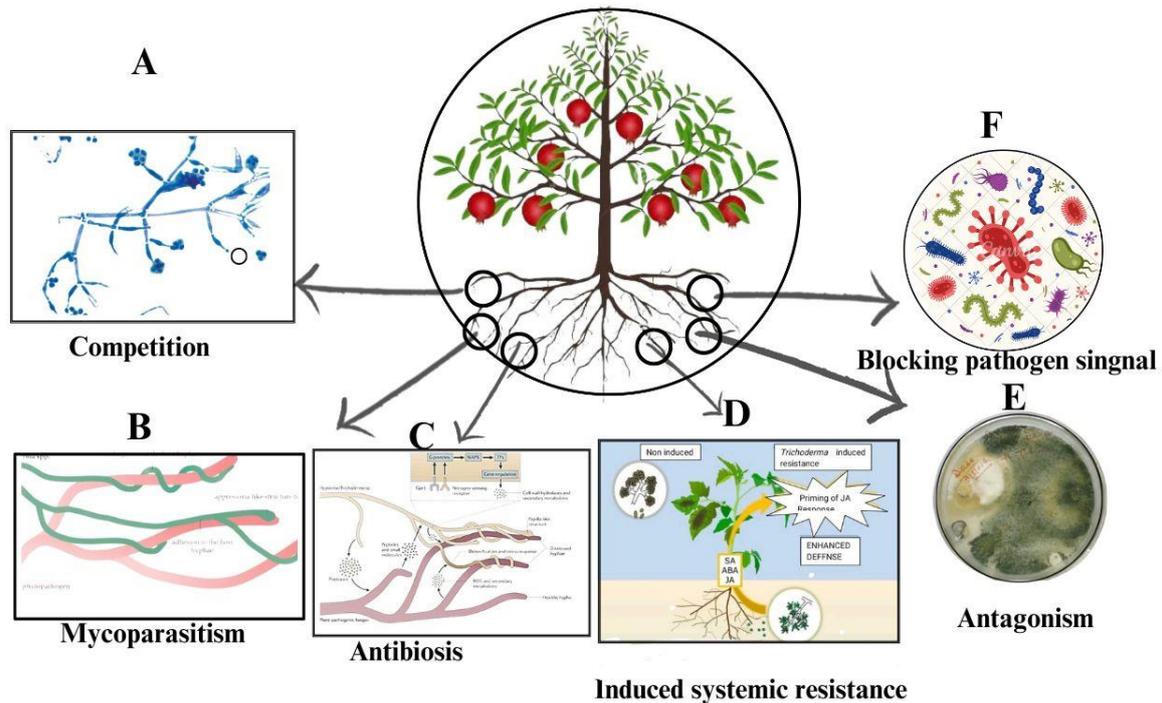


Fig. 1 (A–F). Diagram showing *Trichoderma* mode of action for controlling fungal diseases in plants

3.3.2 Mycoparasitism caused by *Trichoderma*

One of the most important biological controls on *Trichoderma* that exists is mycoparasitism (Fig. 1B). Almost eighteen genera of *Rhizoctonia*, *Phytophthora*, *Peronospora*, and *Pythium* can be parasitized by *Trichoderma*.

3.3.3 Effect of *Trichoderma* antibiosis

It was revealed that antibiosis refers to *Trichoderma*'s capacity to release compounds that are antagonistic to plant pathogenic fungi, hence impeding their growth [33] (Fig. 1C).

3.3.4 *Trichoderma* induction of systemic resistance

Pests may avoid plants that host may *Trichoderma*. Additionally, it can start the process of crops developing defense mechanisms that shield them.

3.3.5 Antagonism with *Trichoderma*

It was explained that the antagonistic effects of *Trichoderma* are thought to arise from the sequential or simultaneous action of many pathways [34] (Fig. 1 –E, F).

3.4 Mycorrhizal fungi

Mycorrhizal fungi have several positive impacts on plant cutting development and survival rates. *Mycorrhizal fungi* are believed to have been widely examined for their impact on horticultural plant developments [35]. Numerous studies have established that *Arbuscular mycorrhizal* symbiosis enhances horticulture yield [36]. *Arbuscular mycorrhizal* fungi (AMF) benefits tree fruit crops by increasing phosphorus absorption and resilience to biotic and abiotic stress, making them more resilient than other plant species. Micropropagation plays a crucial role in bioengineering. Micropropagation is a popular method for producing large quantities of hardwood plants that rely heavily on mycorrhizae. Early mycorrhizal inoculation of micro propagated plantlets improved plant survival [37].

3.5 *Penicillium* spp.

Plant cuttings can be affected by *Penicillium* spp., and these species may function as beneficial endophytes. They improve plant growth and health because of their capacity to colonize plant tissues. The impact of different *Penicillium* species can vary depending on the

type of plant cuttings and environmental conditions. It was revealed that such harmful effects may be reduced by necessary care and by maintaining proper personal cleanliness [44]. Many species of *Penicillium* are harmful to plant cuttings as they stop the cutting from sprouting and developing into a new plant by causing it to degrade and disintegrate [45].

Penicillium spp., sometimes known as moulds, have various beneficial impacts on fruit cuttings. To begin, certain strains of *Penicillium* assist in the natural decomposition procedure through decomposing down biological material and participating in the decomposition of fruit debris, supporting ecologically responsible discarding techniques. Second, some genera of *Penicillium* release catalysts that aid in the breakdown of complex chemicals, which may improve the extraction of flavours and liquids from fruits during cutting [46]. Furthermore, these molds can serve as biocontrol agents, limiting the growth of hazardous pathogens and lowering the risk of spoiling, hence increasing the shelf life of cut fruits. *Penicillium* spp. are also involved in the formation of a variety of useful metabolites, including organic acids and antibacterial substances, which help to preserve fruit quality. However, it is critical to ensure that the strains employed are non-toxic and suitable for eating. With adequate management and awareness, using the beneficial properties of *Penicillium* spp. can result in creative and sustainable procedures in fruit cutting and processing [47].

3.6 *Aspergillus* spp.

It has following kinds of effects on plant cuttings:

3.6.1 Decomposition

These species help break down organic waste in the soil, which might provide nutrients that may be good for the development of plant cuttings [48].

3.6.2 Mycorrhizal associations

Certain *Aspergillus* species have the ability to bind to plant roots and increase the absorption of nutrients and water by the plants [49].

3.6.3 Disease

Damping-off and root rot are two problems that some *Aspergillus* species can produce that are detrimental to plants. Certain diseases might make it difficult for plant cuttings to thrive and survive [50].

3.6.4 Competition

Plant cuttings may not grow and develop as well when they are in competition with one another for nutrients or available space [51].

Aspergillus species can frequently affect plant cuttings in different ways, depending on the environment and the particular species.

Table 2. Examples of bio control of plant pathogens mediated by beneficial fungi

Name of disease	Crop	Causal agent	Biocontrol strain	References
Anthraxnose grey mold	Strawberry (<i>Fragariaananassa</i>)	<i>Colletotrichumacutatum</i> , <i>Botrytis cinerea</i>	<i>T. hamatum</i> <i>T. atroviride</i>	Zin and Badaluddin[38]
Brown root rot	Peanuts(<i>Arachis hypogaea</i>)	<i>Fusariumsolani</i>	<i>T. harzianum</i>	Illa et al, [39]
Root rot disease	Eggplant (<i>Solanum melongena L.</i>)	<i>Macrophominaphaseolina</i>	<i>T. harzianum</i>	Attia et al. [40]
Wilt	Melon (<i>Cucumis melo</i>), Tomato (<i>Solanumlycopersicum</i>)	<i>F. oxysporum</i> and <i>Fusariumoxysporumf.</i> sp. <i>Lycopersici</i>	<i>T.harzianum</i> and T-78 <i>T. asperellum</i>	Boughalleb-M'Hamdi et al.[41]
Fruit rot	Tomato (<i>Solanumlycopersicum</i>)	<i>Rhizoctoniasolani</i>	<i>T. viride</i> , <i>T. virens</i> <i>T. harzianum</i>	Vargas-Inciarte et al.[42] Heflish et al. [43]

Table 3. Beneficial fungi's activities in fruit cultivation

Fruit species	Application effects	References
Banana	Longer branches were observed when treated with AMF and Rhizobacteria.	Ertuark [52]
Strawbery	Reduced levels of malic acid and pH and higher levels of sugar and anthocyanins were observed when treated with AMF and PGPR	Todeschini et al. [53]
Pomegranate	<i>Azotobacterchroococcum</i> or <i>Glomusmosseae</i> combination treatment enhances fruit yield in 5-year-old pomegranate trees.	Naiket al.[54]
Guava	Increased shoot length was obtained when treated with a combination of AMF and <i>Bacillus megaterium</i> .	Emmanuel and Babalola [55]
Citrus	Enhanced growth, improved fruit quality, and improved ion uptake in drought and salinity. Increased soluble sugar and enhanced chlorophyll in foliage as a result of <i>trifoliata mycorrhizal</i> inoculation	Tang et al. [56] Chenget al.[57]
Plum	Plants treated with AMF showed increased vegetative growth.	Pietrantonio et al. [58]

4. INTERACTIONS BETWEEN BACTERIAL AND FUNGALSTRAINS

Positive effects of fruit cuttings depend on the interactions between bacterial and fungalstrains [5]. Fruit crops coexist peacefully with some types of bacteria and fungus. For instance, symbiotic connections between mycorrhizal fungi and plant roots enhance the nutrient-absorbing capacity and general health of the plant. In addition, certain Bacterialstrains could be able to convert ambient nitrogen into a form that plants can use, which would increase the amount of nitrogen available for fruit cuttings. Moreover, certain bacteria create antifungal secondary compounds that shield fruit cuttings from hazardous diseases. The complex network of interactions among these microbes eventually creates an ecology that is healthy and balanced around fruit cuttings, providing ideal circumstances for plant development and fruit yield. Gaining knowledge of and using these microbial interactions might lead to more productive fruit farming and sustainable agricultural systems [59].

5. FUTURE DIRECTION

5.1 Examining particular mechanisms that take place during plant-Bacterial or fungalstrain proliferative interactions [60]. It revealed that additional research is necessary to determine how these bacteria impact signaling molecules, hormones in plants, gene expression, and other aspects of the development and growth of plants.

5.2 An investigation into the possibility of employing certain strains of bacteria and fungi as biocontrol agents in order to enhance the results of plant multiplication.

5.3 Examining the effects of soil composition, temperature, and humidity on the interactions between strains of bacteria, fungi, and plants during propagation. Understanding how these factors impact the beneficial microbes' ability to promote plant growth could lead to more targeted and effective reproduction methods [61].

5.4 An analysis of the effects on plant health and productivity caused by the Bacterial and fungal strains' long-term growth. It might be necessary to conduct fieldwork and monitor plant performance across several growing seasons in order to fully understand the long-term effects of Bacterialinteractions on the development and growth of plants [62].

5.5 Integrating microbial interactions with other plant development management strategies, such as pest control, water management, and nutrient availability [63]. Gaining knowledge of the interactions between various Bacterial and fungal strains and these other elements may lead to more comprehensive and effective plant multiplication promotion strategies [64].

6. CONCLUSION

The investigation of the effects of several bacterial and fungalstrains on fruit cuttings indicates complicated interactions between microbial populations and propagation success.

Compiled literature studies emphasize the importance of individual strains (both Bacterial or fungal) in either supporting or suppressing the formation of roots, focusing on the significance of a more detailed knowledge of microbial relationships in horticulture techniques.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Wahab A, Muhammad M, Munir A, Abdi G, Zaman W, Ayaz A, Khizar C, Reddy SP. Role of arbuscular mycorrhizal fungi in regulating growth enhancing productivity and potentially influencing ecosystems under abiotic and biotic stresses. *Plants*. 2023;12(17):3102.
2. Ghorbanpour M, Omidvari M, Abbaszadeh-Dahaji P, Omidvar R, Kariman K. Mechanisms underlying the protective effects of beneficial fungi against plant diseases. *Biological Control*. 2018;117:147-57.
3. Yadav AN, Kour D, Kaur T, Devi R, Yadav A, Dikilitas M, Abdel-Azeem AM, Ahluwalia AS, Saxena AK. Biodiversity, and biotechnological contribution of beneficial soil microbiomes for nutrient cycling plant growth improvement and nutrient uptake. *Biocatalysis and Agricultural Biotechnology*. 2021;33:102009.
4. Naresh RK, Gupta RK, Panwar AS, Kumar A, Singh B, Dhaliwal SS, Rathore RS, Mahajan NC. Strategies for increasing production potential of horticultural and field crops through improving nutrient-Use efficiency for doubling farming income: A review. *Annals of Horticulture*. 2019;12(1):20-38.
5. Alengebawy A, Abdelkhalek ST, Qureshi SR, Wang MQ. Heavy metals and pesticides toxicity in agricultural soil and plants: Ecological Risks and Human Health Implications. *Toxics*. 2021;9(3):42.
6. Mohsin MZ, Omer R, Huang J, Mohsin A, Guo M, Qian J, Zhuang Y. Advances in engineered *Bacillus subtilis* biofilms and spores, and their applications in bioremediation, biocatalysis, and biomaterials. *Synthetic and Systems Biotechnology*. 2021;6(3):180-91.
7. Hamid B, Zaman M, Farooq S, Fatima S, Sayyed RZ, Baba ZA, Sheikh TA, Reddy MS, El Enshasy H, Gafur A, Suriani NL. Bacterial plant biostimulants: A sustainable way towards improving growth, productivity, and health of crops. *Sustainability*. 2021;13(5):2856.
8. Uzma M, Iqbal A, Hasnain S. Drought tolerance induction and growth promotion by indole acetic acid producing *Pseudomonas aeruginosa* in *Vigna radiata*. *PloS one*. 2022;17(2):e0262932.
9. Worsley SF, Newitt J, Rassbach J, Batey SF, Holmes NA, Murrell JC, Wilkinson B, Hutchings MI. *Streptomyces* endophytes promote host health and enhance growth across plant species. *Applied and Environmental Microbiology*. 2020;86(16):e01053-20.
10. Oleńska E, Małek W, Wójcik M, Swiecicka I, Thijs S, Vangronsveld J. Beneficial features of plant growth-promoting rhizobacteria for improving plant growth and health in challenging conditions: A methodical review. *Science of the Total Environment*. 2020;743:140682.
11. Saeed Q, Xiukang W, Haider FU, Kučerik J, MumtazMZ, Holatko J, Naseem M, Kintl A, Ejaz M, Naveed M, Brtnicky M. Rhizosphere bacteria in plant growth promotion biocontrol, and bioremediation of contaminated sites: A comprehensive review of effects and mechanisms. *International Journal of Molecular Sciences*. 2021;22(19):10529.
12. Bose SK, Howlader P. Melatonin plays a multifunctional role in horticultural crops against environmental stresses: A review. *Environmental and Experimental Botany*. 2020;176:104063.
13. Krieger-Weber S, Heras JM, Suarez C. *Lactobacillus plantarum*, a new biological tool to control malolactic fermentation: A review and an outlook. *Beverages*. 2020;6(2):23.
14. Mofokeng MM, Araya HT, Araya NA, MakgatoMJ, MokgehleSN, Masemola MC, Mudau FN, du Plooy CP, Amoo SO. Integrating biostimulants in agrosystem to promote soil health and plant growth. In *Biostimulants for Crops from Seed Germination to Plant Development*. 2021 ;87-108).
15. Bhardwaj I, Kumar V, Bhardwaj N, Verma R, Bhardwaj Y, Kumari T. A glimpse into the performance and synthesis of microbial nanoparticles and its new advances in soil enrichment and plant nutrition: a review.

- Nanotechnology for Environmental Engineering. 2023;8(4):943-64.
16. Naik K, Mishra S, Srichandan H, Singh PK, Sarangi PK. Plant growth promoting microbes: Potential link to sustainable agriculture and environment. *Biocatalysis and Agricultural Biotechnology*. 2019;21:101326.
 17. Lee H, Yoon Y. Etiological agents implicated in foodborne illness world wide. *Food Science of Animal Resources*. 2021;41(1):1.
 18. Manyi-Loh CE, Mamphweli SN, Meyer EL, Okoh AI. Microbial anaerobic digestion: process dynamics and implications from the renewable energy environmental and agronomy perspectives. *International journal of environmental science and technology*. 2019;16:3913-34.
 19. Firetti R, Martins AN, Suguino E, Turco PH, Araújo HS. Multivariate analysis of vegetative growth and productivity in cultivars of the Cavendish subgroup of the banana. *RevistaCiênciaAgrônômica*. 2023;55:e20228571.
 20. Kurlus R, Rutkowski K, Łysiak GP. Improving of cherry fruit quality and bearing regularity by chemical thinning with fertilizer. *Agronomy*. 2020;10(9):1281.
 21. Khedr E. Improving productivity quality and antioxidant capacity of Le-Conte pear fruits using foliar tryptophan arginine and salicylic applications. *Egyptian Journal of Horticulture*. 2018;45(1):93-103.
 22. Nitnavare R, Bhattacharya J, Ghosh S. Nanoparticles for effective management of salinity stress in plants. *InAgriculturalNanobiotechnology*. WoodheadPublishing.2022;189-216.
 23. Yaseen SA, AL-Zubaydi SR. Effects of foliar application of (nh₄)₂so₄ and alga₂₁st on vegetative growth and chlorophyll content of two cultivars sweet cherry (*Prunus avium L.*) transplants. *Journal of Duhok University*. 2019;22(1):78-88.
 24. Yu YY, Xu JD, Huang TX, Zhong J, Yu H, Qiu JP, Guo JH. Combination of beneficial bacteria improves blueberry production and soil quality. *Food Science & Nutrition*. 2020;8(11):5776-84.
 25. Robledo-Buriticá J, Aristizábal-Loaiza JC, Ceballos-Aguirre N, Cabra-Cendales T. Influence of plant growth-promoting rhizobacteria (PGPR) on blackberry (*Rubus glaucus* Benth. cv. thornless) growth under semi-cover and field conditions. *ActaAgronómica*. 2018;67(2): 258-63.
 26. Bardin M, Gullino ML. Fungal diseases. *Integrated Pest and Disease Management in Greenhouse Crops*. 2020:55-100.
 27. Enyiukwu DN, Basse IN, Nwaogu GA, Chukwu LA, Maranzu JO. Postharvest spoilage and management of fruits and vegetables: A perspective on small-holder agricultural systems of the tropics. *Greener Trends in Plant Pathology and Entomology*. 2020;3:001-17.
 28. Rout S, Tambe S, Deshmukh RK, Mali S, Cruz J, Srivastav PP, Amin PD, Gaikwad KK, de Aguiar Andrade EH, de Oliveira MS. Recent trends in the application of essential oils: The next generation of food preservation and food packaging. *Trends in Food Science & Technology*; 2022.
 29. Bhandari S, Pandey KR, Joshi YR, Lamichhane SK. An overview of multifaceted role of *Trichoderma* spp. for sustainable agriculture. *Archives of Agriculture and Environmental Science*. 2022;6(1):72-9.
 30. Ghazanfar MU, Raza M, Raza W, Qamar MI. *Trichoderma* as potential biocontrol agent, its exploitation in agriculture: a review. *Plant Protection*. 2018;2(3).
 31. Harman G, Khadka R, Doni F, Uphoff N. Benefits to plant health and productivity from enhancing plant microbial symbionts. *Frontiers in Plant Science*. 2021; 11:610065.
 32. Navarro MO, Piva AC, Simionato AS, Spago FR, Modolon F, Emiliano J, Azul AM, Chryssafidis AL, Andrade G. Bioactive compounds produced by biocontrol agents driving plant health. *Microbiome in Plant Health and Disease: Challenges and Opportunities*. 2019:337-74.
 33. Halifu S, Deng X, Song X, Song R, Liang X. Inhibitory mechanism of *Trichoderma virens* ZT05 on *Rhizoctonia solani*. *Plants*. 2020;9(7):912.
 34. Alfiky A, Weisskopf L. Deciphering *Trichoderma*-plant-pathogen interactions for better development of biocontrol applications. *Journal of Fungi*. 2021;7(1):61.
 35. Sui L, Li J, Philp J, Yang K, Wei Y, Li H, Li J, Li L, Ryder M, Toh R, Zhou Y. *Trichoderma atroviride* seed dressing influenced the fungal community and pathogenic fungi in the wheat rhizosphere. *Scientific Reports*. 2022;12(1):9677.

36. Emmanuel OC, Babalola OO. Productivity and quality of horticultural crops through co-inoculation of arbuscular mycorrhizal fungi and plant growth promoting bacteria. *Microbiological Research*. 2020;239:126569.
37. Soumare A, Diédhiou AG, Arora NK, Tawfeeq Al-Ani LK, Ngom M, Fall S, Hafidi M, Ouhdouch Y, Kouisni L, Sy MO. Potential role and utilization of plant growth promoting microbes in plant tissue culture. *Frontiers in Microbiology*. 2021;12:649878.
38. Zin NA, Badaluddin NA. Biological functions of *Trichoderma* spp. for agriculture applications. *Annals of Agricultural Sciences*. 2020;65(2):168-78.
39. Illa C, Torassa M, Pérez MA, Pérez AA. Effect of biocontrol and promotion of peanut growth by inoculating *Trichoderma harzianum* and *Bacillus subtilis* under controlled conditions and field. *Revista Mexicana de Fitopatología*. 2020;38(1):119-31.
40. Attia MS, Hashem AH, Badawy AA, Abdelaziz AM. Biocontrol of early blight disease of eggplant using endophytic *Aspergillus terreus*: improving plant immunological, physiological and antifungal activities. *Botanical Studies*. 2022;63(1):26.
41. Boughalleb-M'Hamdi N, Salem IB, M'Hamdi M. Evaluation of the efficiency of *Trichoderma*, *Penicillium*, and *Aspergillus* species as biological control agents against four soil-borne fungi of melon and watermelon. *Egyptian Journal of Biological Pest Control*. 2018;28:1-2.
42. Vargas-Inciarte L, Fuenmayor-Arrieta Y, Luzardo-Méndez M, Costa-Jardin MD, Vera A, Carmona D, Homen-Pereira M, Costa-Jardin PD, San-Blas E. Use of different *Trichoderma* species in cherry type tomatoes (*Solanum lycopersicum* L.) against *Fusarium oxysporum* wilt in tropical greenhouses. *Agronomía Costarricense*. 2019;43(1):85-100.
43. Heflish AA, Abdelkhalek A, Al-Askar AA, Behiry SI. Protective and curative effects of *Trichoderma asperelloides* Ta41 on tomato root rot caused by *Rhizoctonia solani* Rs33. *Agronomy*. 2021;11(6):1162.
44. Rai S, Singh DK, Kumar A. Microbial, environmental and anthropogenic factors influencing the indoor microbiome of the built environment. *Journal of Basic Microbiology*. 2021;61(4):267-92.
45. Guarnaccia V, Hand FP, Garibaldi A, Gullino ML. Bedding plant production and the challenge of fungal diseases. *Plant Disease*. 2021;105(05):1241-58.
46. de Souza TS, Kawaguti HY. Cellulases, hemicellulases, and pectinases: Applications in the food and beverage industry. *Food and Bioprocess Technology*. 2021;14(8):1446-77.
47. Chilakamarry CR, Sakinah AM, Zularisam AW, Sirohi R, Khilji IA, Ahmad N, Pandey A. Advances in solid-state fermentation for bioconversion of agricultural wastes to value-added products: Opportunities and challenges. *Bioresource technology*. 2022;343:126065.
48. Finlay RD, Thorn RG. The fungi in soil. *Modern Soil Microbiology*, Third Edition. 2019:65-90.
49. Devi R, Kaur T, Kour D, Rana KL, Yadav A, Yadav AN. Beneficial fungal communities from different habitats and their roles in plant growth promotion and soil health. *Microbial Biosystems*. 2020;5(1):21-47.
50. Misra V, Mall AK, Singh D. *Rhizoctonia* Root-Rot Diseases in sugar beet: Pathogen diversity, pathogenesis and cutting-edge advancements in management research. *The Microbe*. 2023 20:100011.
51. Tedersoo L, Bahram M, Zobel M. How mycorrhizal associations drive plant population and community biology. *Science*. 2020;367(6480):eaba1223.
52. ERTÜRK Y. Biological Fertilizers-Containing Beneficial Microorganisms in Fruit Culture. *Kırşehir Ahi Evran Üniversitesi Ziraat Fakültesi Dergisi*. 2022;2(1):71-92.
53. Todeschini V, Ait Lahmidi N, Mazzucco E, Marsano F, Gosetti F, Robotti E, Bona E, Massa N, Bonneau L, Marengo E, Wipf D. Impact of beneficial microorganisms on strawberry growth, fruit production nutritional quality and volatilome. *Frontiers in plant science*. 2018;9:1611.
54. Naik SR, Nandini ML, Jameel Md A, Venkataramana KT, Mukundalakshmi L. Role of Arbuscular Mycorrhiza in Fruit Crops Production. *Int. J. Pure App. Biosci*. 2018;6(5):1126-33.
55. Emmanuel OC, Babalola OO. Productivity and quality of horticultural crops through co-inoculation of arbuscular mycorrhizal fungi and plant growth promoting bacteria.

- Microbiological Research. 2020;239:126569.
56. Tang C, Zhang Z, Yu L, Li Y. Research progress of arbuscular mycorrhizal fungi promoting citrus growth. *Horticulturae*. 2023;9(11):1162.
57. Cheng XF, Wu HH, Zou YN, Wu QS, Kuča K. Mycorrhizal response strategies of trifoliolate orange under well-watered, salt stress, and waterlogging stress by regulating leaf aquaporin expression. *Plant Physiology and Biochemistry*. 2021;162:27-35.
58. Pietrantonio L, Golubkina NA, Cozzolino E, Sellitto M, Cuciniello A, Caruso G. Yield and quality performances of tomatoplum inoculated with arbuscular mycorrhizal fungi in saline soils. InXXX International Horticultural Congress IHC2018: III International Symposium on Innovation and New Technologies in Protected 1271 2018;351-358).
59. Vishwakarma K, Kumar N, Shandilya C, Mohapatra S, Bhayana S, Varma A. Revisiting plant–microbe interactions and microbial consortia application for enhancing sustainable agriculture: A review. *Frontiers in Microbiology*. 2020;11:560406.
60. Fagerlund A, Langsrud S, Møretrø T. Microbial diversity and ecology of biofilms in food industry environments associated with *Listeria monocytogenes* persistence. *Current Opinion in Food Science*. 2021;37:171-8.
61. Arif I, Batool M, Schenk PM. Plant microbiome engineering: expected benefits for improved crop growth and resilience. *Trends in Biotechnology*. 2020;38(12):1385-96.
62. Wang Y, Liu Z, Hao X, Wang Z, Wang Z, Liu S, Tao C, Wang D, Wang B, Shen Z, Shen Q. Biodiversity of the beneficial soil-borne fungi steered by *Trichoderma*-amended biofertilizers stimulates plant production. *NPJ Biofilms and Microbiomes*. 2023;9(1):46.
63. Umesha S, Singh PK, Singh RP. Microbial biotechnology and sustainable agriculture. In *Biotechnology for Sustainable Agriculture* 2018;185-205).
64. Kokkoris V, Hart M. *In vitro* propagation of arbuscular mycorrhizal fungi may drive fungal evolution. *Frontiers in Microbiology*. 2019;10:2420.

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